

Transamination Reaction of Hydrophobic Pyridoxal with an α -Amino Acid in Functionalized Bilayer Vesicles: Co-operative Catalysis by the Imidazolyl Group and Copper(II) Ions

Yukito Murakami,* Jun-ichi Kikuchi, Toru Imori, and Kazunari Akiyoshi

Department of Organic Synthesis, Faculty of Engineering, Kyushu University, Fukuoka 812, Japan

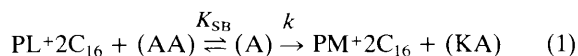
1-(*N,N*-Dihexadecylcarbamoylmethyl)-2-methyl-3-hydroxy-4-formyl-5-hydroxymethylpyridinium chloride (PL+2C₁₆) undergoes a transamination reaction with *L*-phenylalanine in single-walled bilayer vesicles formed from two different peptide lipids (N⁺C₅Ala2C₁₆ and N⁺C₅His2C₁₆); co-ordination of copper(II) ion to the Schiff-base intermediate results in a marked rate acceleration.

The aim of our research is to create holoenzyme models to simulate some functional aspects of vitamin B₆-dependent enzymes. We have based our work on the following information: (i) that Schiff-base formation between pyridoxal and a hydrophobic α -amino acid is much enhanced in molecular aggregates such as micelles and vesicles, so that the overall transamination reaction is considerably accelerated;¹⁻³ (ii) a positive charge on the pyridyl nitrogen of pyridoxal provides an electron sink which promotes the α -hydrogen abstraction from an aldimine Schiff-base;^{2,4} (iii) when the pyridoxal moiety is fixed in the so-called hydrogen-belt domain formed by the histidyl residues interposed between the hydrophobic and hydrophilic zones within the vesicles, the imidazolyl group induces an intramolecular prototropic shift to give the corresponding ketimine Schiff-base and accelerates the overall transamination reaction.^{1,3} We report here on the transamination reaction of PL+2C₁₆† with *L*-phenylalanine (*L*-Phe) (Scheme 1) in single-walled bilayer vesicles formed from the synthetic peptide lipids, N⁺C₅Ala2C₁₆‡ and N⁺C₅His2C₁₆‡.

Aqueous dispersions of either of the peptide lipids (1.0 \times 10⁻³ mol dm⁻³) containing PL+2C₁₆ (5.0 \times 10⁻⁵ mol dm⁻³) in an aqueous carbonate buffer (2.0 \times 10⁻² mol dm⁻³, pH 9.9, μ 0.10 M with KCl) were sonicated for 1 min with a probe-type sonicator at 30 W to give single-walled bilayer vesicles. The transamination reaction in the vesicles was followed spectrophotometrically at 30.0 °C. When PL+2C₁₆ is incorporated into the respective vesicles formed from N⁺C₅Ala2C₁₆ and

N⁺C₅His2C₁₆, these vesicles provide for the pyridoxal moiety a microenvironment equivalent to propan-1-ol in polarity [*E*_T(30) = 50.7]. The extent of formation of an aldimine Schiff-base of PL+2C₁₆ with an α -amino acid in the vesicles is dependent on the hydrophobicity of the α -amino acid (equilibrium constants *K*_{SB} at pH 9.9 in the N⁺C₅Ala2C₁₆ vesicle: *L*-Ala ca. 10, *L*-Leu 560, *L*-Phe 1700 mol⁻¹ dm³).

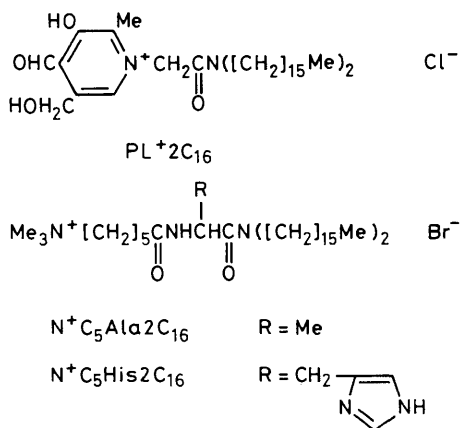
As observed for the transamination reaction of pyridoxal-5'-phosphate with *N*-dodecyl-*L*-alaninamide in vesicles,¹ the reaction proceeded through the fast equilibrated formation of an aldimine Schiff-base [(A) in Scheme 1] followed by much slower conversion into the pyridoxamine derivative (PM+2C₁₆) and β -phenylpyruvate (KA).§ The rate-determining step in the overall reaction is the isomerization of the aldimine Schiff-base [(A) in Scheme 1] to the corresponding ketimine [(C) in Scheme 1] since the accumulation of the ketimine Schiff-base was not observed to any detectable extent during the reaction. The pH-dependent equilibrium constants (*K*_{SB}) for the formation of (A) and the rate constants (*k*) for the isomerization, as defined in equation (1) [(AA) for *L*-Phe], are listed in Table 1.¶



The *K*_{SB} value for 1,2-dimethyl-3-hydroxy-4-formyl-5-hydroxymethylpyridinium chloride⁵ with *L*-Phe is extremely low (10 mol⁻¹ dm³) in aqueous carbonate buffer (2.0 \times 10⁻² mol dm⁻³, pH 9.9, μ 0.10 M with KCl) at 30.0 °C. The transamination was followed spectrophotometrically in the presence of a 1000-fold excess of *L*-Phe (5.0 \times 10⁻² mol dm⁻³) over the pyridoxal derivative; *k*_{obs} = 2 \times 10⁻⁶, *k* = 6 \times 10⁻⁶ s⁻¹. Since this *k* value is comparable with those obtained for the vesicular systems, the overall rate acceleration in the vesicles originates predominantly from the enhancement of Schiff-base formation.

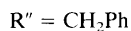
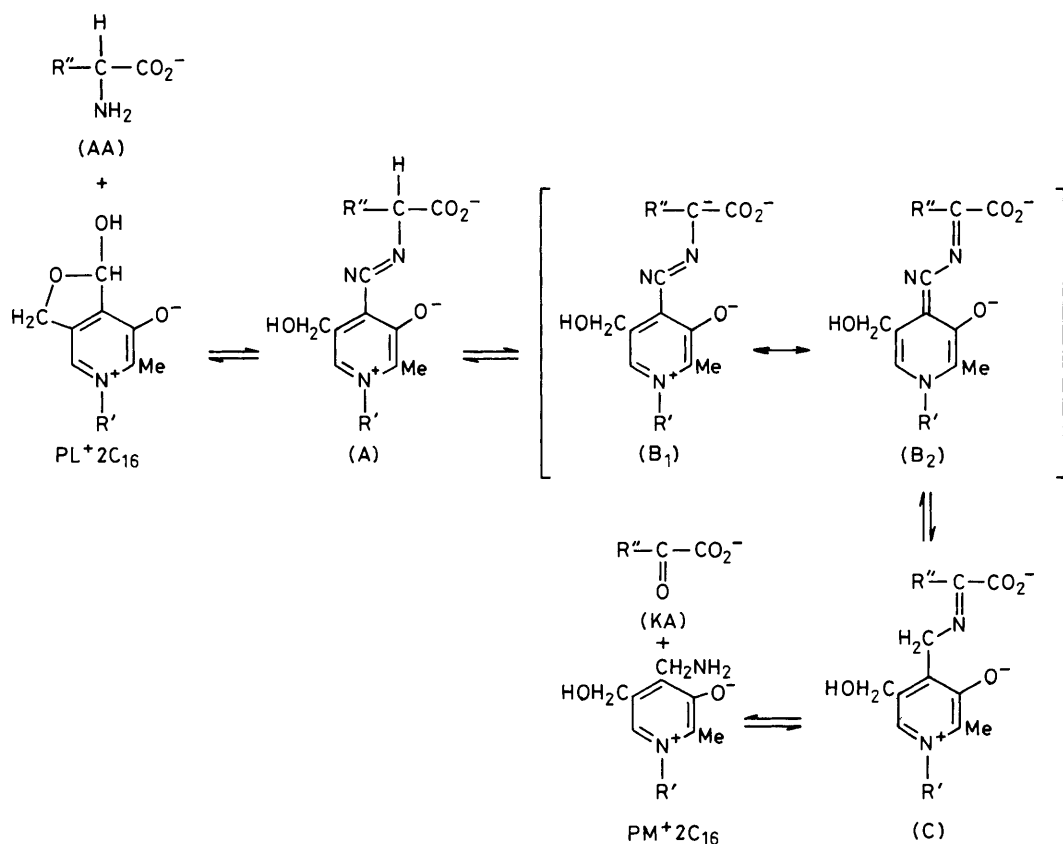
§ PL+2C₁₆ was separated from the reaction mixture by extraction with chloroform. The extracted PL+2C₁₆ and the (KA) remaining in the reaction medium were separately converted into the corresponding 2,4-dinitrophenylhydrazones and their amounts were quantitatively evaluated by spectrophotometric means. PM+2C₁₆ was extracted with toluene from the reaction mixture and then treated with 2,4,6-trinitrobenzenesulphonate before identification and quantitative evaluation by spectrophotometry: A. P. H. Phan, T. T. Ngo, and H. M. Lenhoff, *Appl. Biochem. Biotech.*, 1983, **8**, 127. When the transamination was completed both in the presence and absence of copper(II) ions, PL+2C₁₆ was not detected in the reaction mixture and PM+2C₁₆ and (KA) were obtained quantitatively.

¶ The apparent first-order rate constant (*k*_{obs}) for the disappearance of the aldimine Schiff-base was calculated from the initial rate of absorbance decay at 393 nm and is correlated with *K*_{SB} and *k* by the equation: *k*_{obs} = *kK*_{SB}[AA]₀/(*K*_{SB}[AA]₀ + 1); where [AA]₀ denotes the initial concentration of the amino acid (ref. 2).



† PL+2C₁₆ was prepared by the reaction of pyridoxal monomethylacetal (ref. 5) with *N,N*-dihexadecyl-2-iodoacetamide, followed by hydrolysis to give a yellow solid, characterised by elemental analysis and n.m.r. spectroscopy.

‡ N⁺C₅His2C₁₆ was prepared as described previously (ref. 1) as a colourless glassy solid, characterised by elemental analysis and n.m.r. spectroscopy.



Scheme 1

Addition of copper(II) ions to the reaction system in equation (1) caused a drastic change in reaction behaviour. When copper(II) perchlorate (2-fold molar excess over PL+2C₁₆) was added to the equilibrium mixture of the aldimine Schiff-base, PL+2C₁₆, and L-Phe, obtained in the initial fast step, the spectral changes shown in (3) in Figure 1 were observed in both vesicular systems. Two different Cu^{II} complex species, which have absorption maxima at 523 and *ca.* 390 nm and are mutually in equilibrium, were formed within 5 s,** and the former species underwent reaction to give PM+2C₁₆ and (KA).

The 523 nm band that appeared upon addition of copper(II) ions is due to the copper(II) complex of the carbanion

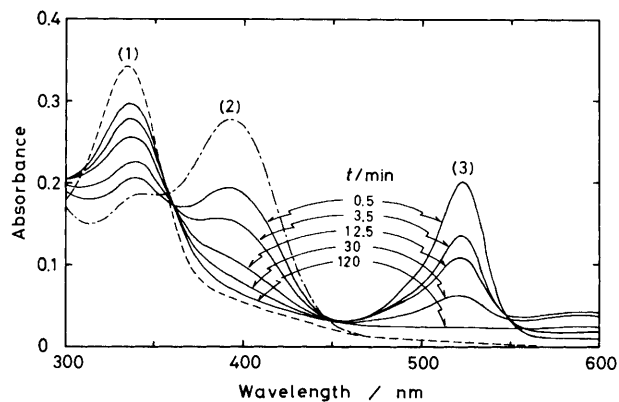


Figure 1. Absorption spectral changes for the Cu^{II}-catalysed transamination reaction of PL+2C₁₆ (5.0×10^{-5} mol dm⁻³) with L-Phe (5.0×10^{-3} mol dm⁻³) in the presence of N⁺C₅Ala2C₁₆ (1.0×10^{-3} mol dm⁻³) in aqueous carbonate buffer (2.0×10^{-2} mol dm⁻³, pH 9.9, μ 0.10 M with KCl) at 30.0 °C: (1) (---) PL+2C₁₆ alone; (2) (-·-·-) equilibrium mixture of PL+2C₁₆, L-Phe, and (A) (Scheme 1) without Cu^{II}; (3) (—) Cu^{II} complex species formed upon addition of Cu(ClO₄)₂ (1.0×10^{-4} mol dm⁻³) to (2). The curves in (3) are distinguished by figures which give the reaction period (*t*) in min after addition of copper(II) ions.

** The absorption maximum at *ca.* 390 nm for the Cu^{II}-(A) complex, which was formed instantaneously upon addition of copper(II) perchlorate to (A), is very close to that for (A). PL+2C₁₆ (5.0×10^{-5} mol dm⁻³) underwent reaction with L-Phe (5.0×10^{-3} mol dm⁻³) to afford the corresponding aldimine Schiff-base at *ca.* 30% conversion at pH 7.6 in the N⁺C₅Ala2C₁₆ vesicle. The absorption band at *ca.* 400 nm increased instantaneously upon addition of copper(II) perchlorate (1.0×10^{-4} mol dm⁻³) without showing any significant band shift. The copper(II) complex of the aldimine Schiff-base formed from 2-methyl-3-hydroxy-4-formyl-5-dodecylthiomethylpyridine and L-alanine shows an absorption maximum at *ca.* 390 nm in vesicular systems as reported previously (ref. 3).

Table 1. Kinetic parameters for the reaction of PL+2C₁₆ with L-Phe in single-walled vesicles at 30.0 °C.^a

Peptide lipid	Metal-free ^b			Cu ^{II} -catalysed ^c $k_{\text{obs}}^{\text{Cu}} \times 10^3/\text{s}^{-1}$
	$k_{\text{obs}} \times 10^6/\text{s}^{-1}$	$K_{\text{SB}}/\text{mol}^{-1} \text{ dm}^3$	$k \times 10^6/\text{s}^{-1}$	
N+C ₅ His2C ₁₆	6.5	830	8.1	23
N+C ₅ Ala2C ₁₆	1.5	1700	1.6	4.3

^a Initial concentrations in mol dm⁻³: PL+2C₁₆, 5.0×10^{-5} ; L-Phe, 5.0×10^{-3} ; peptide lipids, 1.0×10^{-3} . Medium: 2.0×10^{-2} mol dm⁻³ aqueous carbonate buffer (pH 9.9) containing 0.10 mol dm⁻³ KCl. ^b Ethylenediaminetetra-acetic acid (1.0×10^{-4} mol dm⁻³) was added. ^c Cu(ClO₄)₂ (1.0×10^{-4} mol dm⁻³) was added immediately after aldimine Schiff-base formation.

intermediate [(B₁) and (B₂) in Scheme 1] derived by α-hydrogen abstraction from the Cu^{II}-aldimine Schiff-base chelate. This is based on the following observations. (i) When DL-phenylglycine was substituted for L-Phe, the corresponding absorption band appeared at 533 nm (red shift by 10 nm), whereas the combination of PL+2C₁₆ and dodecylamine did not show such a spectral change upon addition of copper(II) ions. (ii) Addition of 5,5'-dithiobis(2-nitrobenzoic acid) which reacts with carbanions, to the copper(II)-containing system gave instantaneously a new absorption band at ca. 420 nm, a characteristic band for the 3-carboxy-4-nitrothiophenolate anion, and caused the disappearance of the 523 nm band. (iii) When methyl L-phenylalaninate was used as a substrate, a species with an absorption maximum at 502 nm, a characteristic band for the carbanion intermediate, was formed even in the absence of copper(II) ions. Upon addition of copper(II) perchlorate, the band was shifted to 523 nm. (iv) Such carbanion intermediates have been detected in enzymic and nonenzymic systems,⁷ although there is very limited formation of the carbanion intermediate in nonenzymic systems containing α-amino acids. ||

In conclusion, the co-ordination of copper(II) ion to the aldimine Schiff-base significantly enhanced deprotonation at the α-carbon atom so that the rate-determining step was

changed from (A)→(C) to (B₁), (B₂)→(C) (Scheme 1) and the isomerization was accelerated dramatically (Table 1). The higher reactivity in the N+C₅His2C₁₆ vesicle relative to that in the N+C₅Ala2C₁₆ vesicle must originate from the catalytic function of the imidazolyl group. The single-walled vesicles formed from peptide lipids can provide extremely effective reaction sites for acceleration of transamination reactions.

Received, 11th June 1984; Com. 806

References

- 1 Y. Murakami, A. Nakano, and K. Akiyoshi, *Bull. Chem. Soc. Jpn.*, 1982, **55**, 3004.
- 2 H. Kondo, J. Kikuchi, and J. Sunamoto, *Tetrahedron Lett.*, 1983, 2403.
- 3 Y. Murakami, J. Kikuchi, A. Nakano, K. Akiyoshi, and T. Imori, *Bull. Chem. Soc. Jpn.*, 1984, **57**, 1116.
- 4 J. R. Maley and T. C. Bruice, *Arch. Biochem. Biophys.*, 1970, **136**, 187.
- 5 D. Heyl, E. Luz, S. A. Harris, and K. Folkers, *J. Am. Chem. Soc.*, 1951, **73**, 3430.
- 6 Y. Murakami, A. Nakano, A. Yoshimatsu, K. Uchitomi, and Y. Matsuda, *J. Am. Chem. Soc.*, 1984, **106**, 3613.
- 7 W. T. Jenkins, *J. Biol. Chem.*, 1964, **239**, 1742; F. S. Furbish, M. L. Fonda, and D. E. Metzler, *Biochemistry*, 1969, **8**, 5169; M. Martinez-Carrion, D. C. Tiemeier, and D. L. Peterson, *ibid.*, 1970, **9**, 2574; L. Schirch and R. A. Slotter, *ibid.*, 1966, **5**, 3175; J. R. Maley and T. C. Bruice, *J. Am. Chem. Soc.*, 1968, **90**, 2843; S. Matsumoto and Y. Matsushima, *ibid.*, 1972, **94**, 7211; S. Matsumoto and Y. Matsushima, *ibid.*, 1974, **96**, 5228.

|| The carbanion intermediate was not detected during the reaction of 1,2-dimethyl-3-hydroxy-4-formyl-5-hydroxymethylpyridinium chloride with L-Phe in the presence of copper(II) ion in aqueous carbonate buffer at pH 9.9, μ 0.10 M, and 30.0 °C.